

Circulating CLA⁺ T cell effector function allows patient stratification for IL-13 and IL-31 production in adult non-treated moderate-to-severe atopic dermatitis

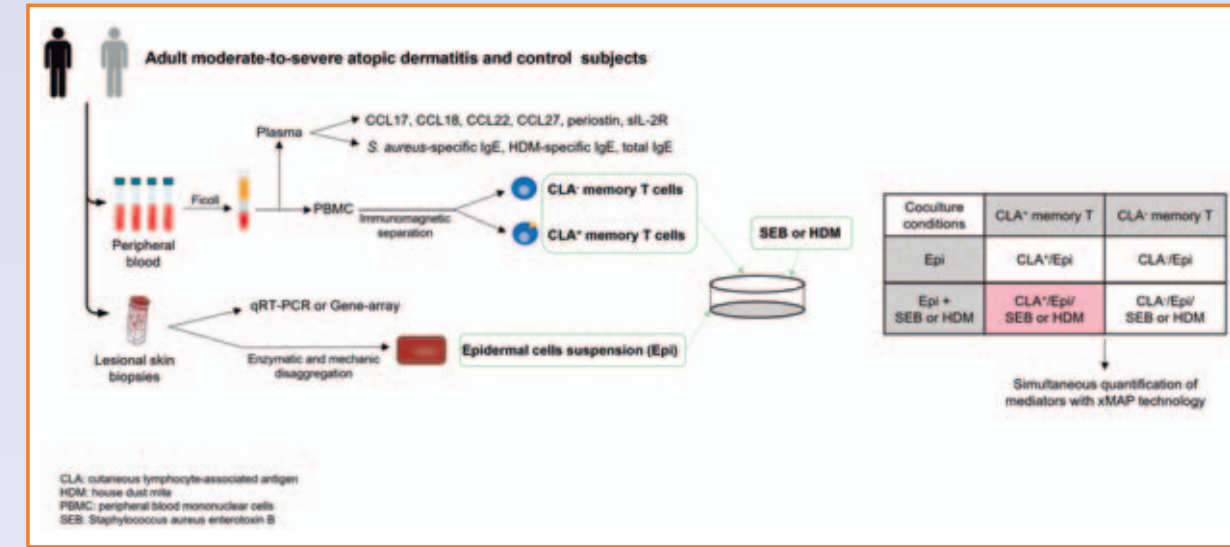
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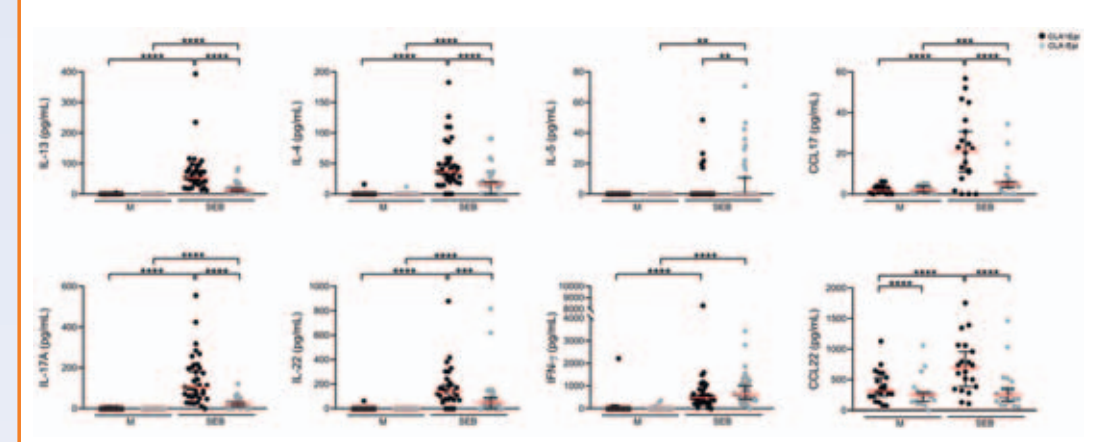
Background

Atopic dermatitis (AD) is a chronic heterogeneous T-cell mediated skin disease. Circulating skin-homing cutaneous lymphocyte-associated antigen (CLA)⁺ memory T cells reflect the cutaneous abnormalities in AD^{1,2}. There is currently no clear *in vitro* functional immunological approach to explore endotypes for Th2 cytokines. *Staphylococcus aureus*, CLA⁺ memory T cells and IL-13 are relevant players in AD. *S. aureus* colonizes AD skin and is associated to flare ups, disease severity and enhanced Th2 immune response. Application of *S. aureus* enterotoxin B (SEB) to intact AD skin induces T cells activation. We aimed at understanding AD functional immune Th2 response heterogeneity through SEB activation of CLA⁺ memory T cells. IL-31 is produced by dendritic cells, mast cells, basophils, eosinophils, type 2 innate lymphoid cells, M2 macrophages, and preferentially CD4⁺ T cells with a Th2 phenotype. It is involved in inflammation, pruritus, epidermal barrier disruption and tissue remodelling. The relationship between IL-31 and allergens has been poorly characterized and mainly limited to canine models. IL-31 production by T cells is mostly studied through qRT-PCR and intracellular flow cytometry of peripheral blood mononuclear cells in a low number of patients, and without association with the clinic. We aimed at deciphering the role of allergen in IL-31 production by memory T cells and its possible relationship with clinical features in AD patients.

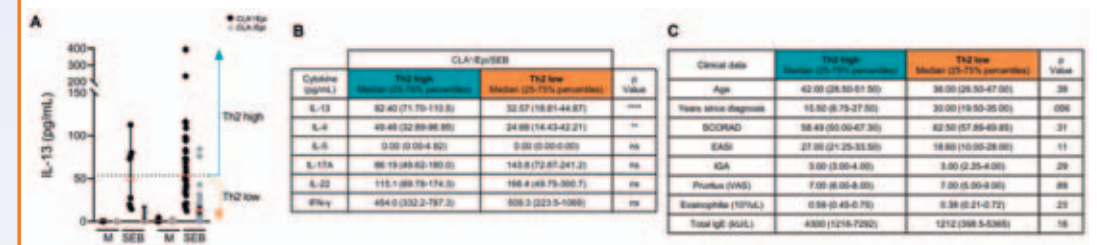
Methods



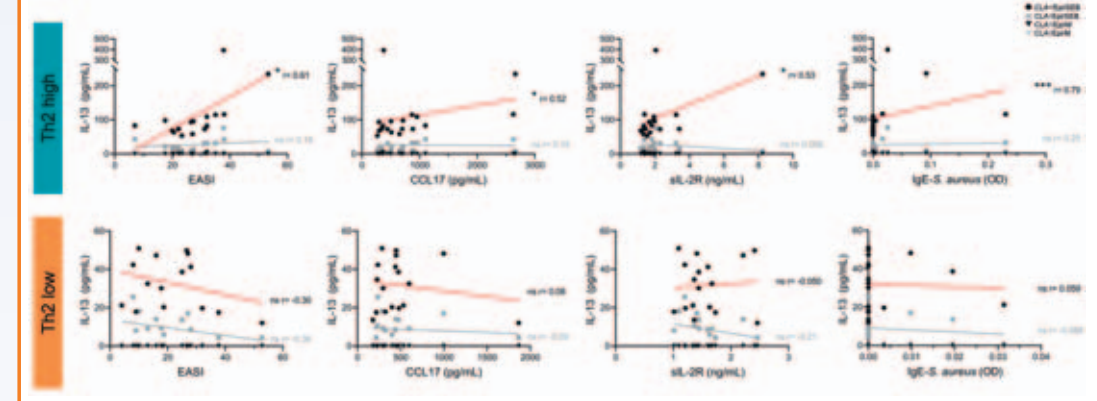
SEB-induced IL-13 production in CLA⁺ memory T cells defines Th2 high and Th2 low responders in atopic dermatitis³



IL-13, IL-4, IL-17A, IL-22, CCL17 and CCL22 are preferentially produced by circulating CLA⁺ memory T cells activated with SEB. Quantification (pg/mL) of IL-13, IL-4, IL-5, IL-17A, IL-22, IFN-γ, CCL17, and CCL22 in 24-hour cocultures in basal conditions (M) or stimulated with SEB (n = 35 for IL-13/4/17A and IFN-γ, n = 30 for IL-5, n = 29 for IL-22, and n = 20 for CCL17/22). Statistics are represented as median with 95% confidence interval. Wilcoxon test was used for comparisons. **: p<.01; ***: p<.001; ****: p<.0001.

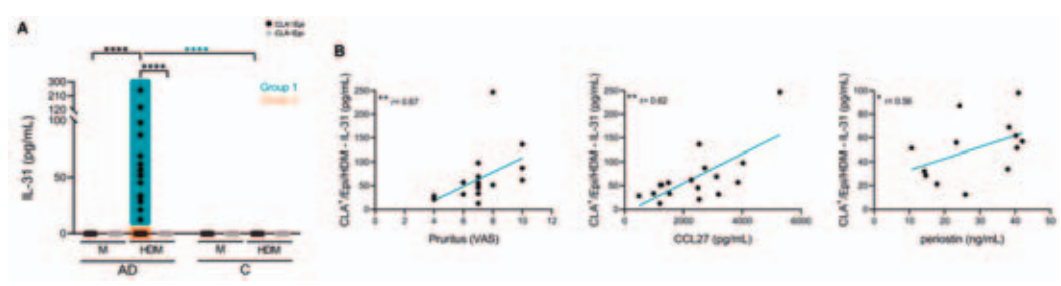


The IL-13 response enables the stratification of a clinically homogeneous population into Th2 high and Th2 low groups. (A) IL-13 levels (pg/mL) in AD (n = 35) and control subjects (C, n = 8). Dotted line indicates the median of SEB-induced CLA⁺ T-cell IL-13 response in AD. (B) IL-13, IL-4, IL-5, IL-17A, IL-22 and IFN-γ (pg/mL) produced by SEB-induced CLA⁺ T-cell cocultures was compared between Th2 high (n = 14-18) and Th2 low (n = 15-17) groups. Data presented as the median (25-75% percentiles) and compared with the Mann-Whitney test. (C) Clinical characteristics were compared between Th2 high (n = 12-17) and Th2 low (n = 13-17) groups. Data presented as the median (25-75% percentiles) and compared with the Mann-Whitney test. EASI, eczema area and severity index; IGA, investigator's global assessment; M, untreated; SCORAD, scoring atopic dermatitis; VAS, visual analogue scale. ns: p>.05; **: p<.01; ****: p<.0001.



In the Th2 high group, unlike the Th2 low, the SEB-induced CLA⁺ T-cell IL-13 response directly correlates with eczema area and severity index (EASI), CCL17, sIL-2R and *S. aureus*-specific IgE plasma levels. IL-13 (pg/mL) from 24-hour cocultures was correlated with EASI (n = 17 for Th2 high and n = 15 for Th2 low), and plasma levels of CCL17 (n = 18 for Th2 high and n = 17 for Th2 low), sIL-2R (n = 18 for Th2 high and n = 17 for Th2 low), and *S. aureus*-specific IgE (n = 17 for Th2 high and n = 17 for Th2 low). Correlations performed with R Spearman test and represented with linear regression. M, untreated; ns: p>.05; *: p<.05; ***: p<.001.

Allergen sensitization stratifies IL-31 production by CLA⁺ memory T cells in atopic dermatitis⁴

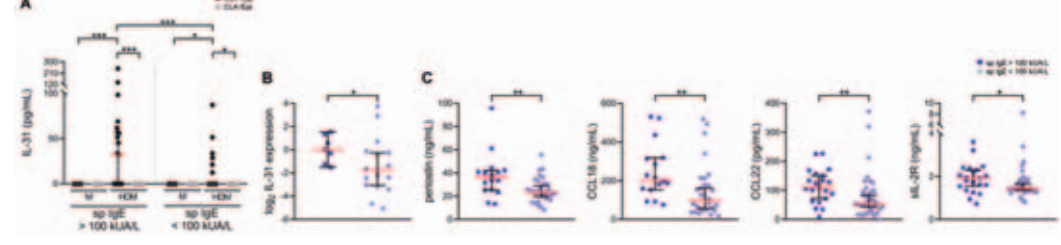


HDM induces IL-31 in CLA⁺ memory T cells cultured with autologous lesional epidermal atopic dermatitis cells, which correlates with pruritus, and plasma CCL27 and periostin. (A) IL-31 (pg/mL) produced by CLA⁺ memory T-cell cocultures in basal conditions (M) or stimulated with HDM in AD (n = 58) and C- (n = 11) derived samples. Statistics are represented as median with 95% confidence interval. Wilcoxon test and Mann-Whitney test were used for comparisons within the same group or between two groups, respectively. (B) Correlations of CLA⁺ Epi/HDM IL-31 response in group 1 with pruritus (n = 17), and plasma levels of CCL27 (n = 17), and periostin (n = 13). Correlations tested with R Spearman and represented with linear regression. C, control subjects; VAS, visual analogue scale. *: p<.05; **: p<.01; ****: p<.0001.

A			
Clinical data	Group 1	Group 2	P Value
Age	28.00 (18.50-44.00)	36.00 (27.00-45.75)	.063
Years since diagnosis	15.50 (11.00-25.75)	24.00 (13.00-33.50)	.28
SCORAD	59.63 (36.00-61.32)	59.27 (42.10-68.00)	.44
EASI	22.45 (19.38-27.93)	25.00 (15.25-32.00)	.32
IGA	4.00 (3.00-4.00)	3.00 (3.00-4.00)	.35
Pruritus (VAS)	7.00 (6.50-8.00)	8.00 (6.25-9.00)	.31
Eosinophils (10 ⁹ /L)	0.73 (0.21-1.06)	0.48 (0.24-0.81)	.56
sp IgE response (OD)	187.47 (123.87-284.56)	533.6 (283.9-982.9)	<.0001
Total IgE (kU/L)	2000 (889.9-5080)	588.6 (116.5-1792)	.0027

B			
Cytokine (pg/mL)	Group 1	Group 2	P Value
IL-13	242.7 (123.7-471.8)	5.22 (0.00-41.24)	****
IL-4	57.34 (32.55-103.5)	0.00 (0.00-6.53)	****
IL-5	67.72 (12.04-138.6)	0.00 (0.00-6.71)	****
IL-17A	24.83 (9.98-49.25)	0.00 (0.00-7.75)	***
IL-22	57.18 (11.16-149.0)	0.00 (0.00-0.00)	****
IFN-γ	0.00 (0.00-22.97)	0.00 (0.00-7.32)	ns

Patients with IL-31 response by HDM-activated CLA⁺ T cells show higher HDM-specific and total IgE and more inflammatory profile than patients with no IL-31 response. (A) Clinical characteristics were compared between group 1 (n = 15-17) and group 2 (n = 31-41) of patients. Data presented as the median (25-75% percentiles) and compared with the Mann-Whitney test. Bold values indicate significant data. (B) IL-13, IL-4, IL-5, IL-17A, IL-22 and IFN-γ (pg/mL) produced by HDM-induced CLA⁺ T-cell cocultures were compared between groups 1 (n = 13-17) and 2 (n = 35-41) of patients. Data presented as the median (25-75% percentiles) and compared with the Mann-Whitney test. (C) Functional enrichment of all differential expressed genes (gene-array performed in n = 3 group 1 and n = 13 group 2 cutaneous lesions, considering differential expressed genes those with a fold change of 1.5 or greater and a p value of less than .05) up-regulated in group 1 and group 2 of patients with GO biological process. For group 1, summary of enriched biological processes; for group 2, representation of all enriched biological processes of significant terms (Benjamini-Hochberg FDR < .05). EASI, eczema area and severity index; IGA, investigator's global assessment; GO, gene ontology; OD, optical density; SCORAD, scoring atopic dermatitis; sp IgE, HDM-specific IgE; VAS, visual analogue scale. ns: p>.05; ***: p<.001; ****: p<.0001.



Patients with HDM-specific IgE > 100 kU/L show increased IL-31 response by HDM-induced CLA⁺ T cells, IL-31 mRNA expression in lesional skin and plasma levels of periostin, CCL18, CCL22 and sIL-2R compared to those with HDM-specific IgE < 100 kU/L. (A) IL-31 (pg/mL) from atopic dermatitis culture supernatants separated according to sp IgE levels (> 100 kU/L, n = 21, < 100 kU/L, n = 37). Statistics are represented as median with 95% confidence interval, and Wilcoxon test and Mann-Whitney test were used for comparisons within the same group or between two groups, respectively. (B) IL-31 mRNA expression in cutaneous lesions (high n = 7, low n = 17) and (C) plasma levels of periostin (ng/mL), CCL18 (ng/mL), CCL22 (pg/mL) and sIL-2R (ng/mL) were compared between high (n = 16-21) and low (n = 30-37) sp IgE groups. Statistics are represented as median with 95% confidence interval, Mann-Whitney test was used for comparisons. M, untreated; sp IgE, HDM-specific IgE. *: p<.05; **: p<.01; ***: p<.001.

Conclusions

This translational approach based on the coculture of skin-homing CLA⁺ memory T cells with autologous lesional epidermal cells from adult non-treated moderate-to-severe atopic dermatitis patients allows identifying Th2 high and Th2 low responders from a clinically homogeneous population based on the SEB-CLA⁺IL-13 axis that differently correlate with EASI and CCL17, the best biomarker for atopic dermatitis. Additionally, it allows stratifying patients into IL-31 producers and non-producers by HDM-induced CLA⁺ T cells in relation with allergen sensitization status by functionally analysing the CLA⁺ T-cell immune response, and correlating IL-31 response with patient's pruritus and CCL27, a biomarker of clinical response to anti-IL-31RA treatment. The current findings may help to explore the complex heterogeneity of AD pathophysiology from a more functional point of view and guide patient's stratification for directed therapies.